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(54) Title: HYBRID WITH INTERFERON- α AND AN IMMUNOGLOBULIN Fc LINKED THROUGH A NON-IMMUNOGENIC PEPTIDE		
(57) Abstract		
Disclosed is a hybrid recombinant protein consisting of human interferon, preferably interferon- α (IFN α), and human immunoglobulin Fc fragment, preferably γ 4 chain, joined by a peptide linker comprising the sequence Gly Gly Ser Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser (SEQ ID NO:1).		

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**Hybrid with Interferon- α and an Immunoglobulin Fc
Linked through a Non-immunogenic Peptide**

5 **Background of the invention**

Interferon- α ("IFN α ") was among the first of the cytokines to be produced by recombinant DNA technology and has been shown to have therapeutic value in conditions such as inflammatory, viral, and malignant diseases. Several IFN α preparations, including those purified from the natural sources and those generated by recombinant DNA technology, have been used or are being tested in a variety of malignant and viral diseases. IFN α can cause regression of some established tumors and induce positive responses in some viral infections. So far, IFN α has been approved or tested in many countries for indications such as: Kaposi's sarcoma; hairy cell leukemia; malignant melanoma; basal cell carcinoma; multiple myeloma; renal cell carcinoma, hepatitis B; hepatitis C; venereal warts, Herpes I/II, varicella/herpes zoster; and mycosis fungoides.

Most cytokines, including IFN α , have relatively short circulation half-lives since they are produced *in vivo* to act locally and transiently. The serum half-life of IFN α is only about two to eight hours (Roche Labs. Referon A, Schering Intron A, *Physicians' Desk Reference*, 47th edition, 1993, pp. 2006-2008, 2194-2201). To use IFN α as an effective systemic therapeutic, one needs relatively large doses and frequent administrations. For example, one of the recommended therapeutic strategies for the AIDS-related Kaposi's sarcoma starts with an induction dose of 36 million IU daily for 10 to 12 weeks, administered as an intramuscular or subcutaneous injection, followed by a maintenance dose of 36 million IU, three times a week. (Roche Labs. Referon A, *Physicians' Desk Reference*, 47th edition, 1993, pp. 2006-2008). Such frequent parenteral administrations are inconvenient and painful. Further, toxic effects, which are probably caused by the high dosage, are a problem for certain

patients. Skin, neurologic, endocrine, and immune toxicity have been reported. To overcome these disadvantages, one can modify the molecule to increase its circulation half-life or change the drug's formulation to extend its release time. The dosage and administration frequency can then be reduced while increasing the efficacy. It was reported that doses of less than nine million units had been well tolerated, while doses more than 36 million units can induce severe toxicity and significantly alter patient status. (Quesada, J.R. et al., *J. Clin. Oncol.*, 4:234-43, 1986). It is possible to decrease substantially the toxic effects by producing a new form IFN α which is more stable in the circulation and requires smaller doses. Efforts have been made to create a recombinant IFN α -gelatin conjugate with an extended retention time (Tabata, Y. et al., *Cancer Res.* 51:5532-8, 1991). A lipid-based encapsulated IFN α formulation has also been tested in animals and achieved an extended release of the protein in the peritoneum (Bonetti, A. and Kim, S. *Cancer Chemother Pharmacol.* 33:258-261, 1993).

Immunoglobulins of IgG and IgM class are among the most abundant proteins in the human blood. They circulate with half-lives ranging from several days to 21 days. IgG has been found to increase the half-lives of several ligand binding proteins (receptors) when used to form recombinant hybrids, including the soluble CD4 molecule, LHR, and IFN- γ receptor (Mordini J. et al., *Nature*, 337:525-31, 1989; Capon, D.J. and Lasky, L.A., U.S. Patent number 5,116,964; Kurschner, C. et al., *J. Immunol.* 149:4096-4100, 1992). However, such hybrids can present problems in that the peptide at the C-terminal of the active moiety and the peptide at the N-terminal of the Fc portion at the fusion point creates a new peptide sequence, which is a neoantigen, and which can be immunogenic. The invention relates to a IFN α -Fc hybrid which is designed to overcome this problem and extend the half-life of the IFN α .

Summary of the invention

The present invention relates to a hybrid recombinant protein which consists of two subunits. Each subunit includes a human interferon, preferably IFN α , joined by a peptide linker which is primarily composed of a T cell inert sequence, linked to a human immunoglobulin Fc fragment, preferably the $\gamma 4$ chain. The $\gamma 4$ chain is preferred over the $\gamma 1$ chain because the former has little or no complement activating ability.

The C-terminal end of the IFN α is linked to the N-terminal end of the Fc fragment. An additional IFN α (or other cytokine) can attach to the N-terminal end of any other unbound Fc chains in the Fc fragment, resulting in a homodimer for the $\gamma 4$ chain. If the Fc fragment selected is another chain, such as the μ chain, then, because the Fc fragments form pentamers with ten possible binding sites, this results in a molecule with interferon or other cytokine linked at each of ten binding sites.

The two moieties of the hybrid are linked through a T cell immunologically inert peptide (e.g., Gly Gly Ser Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser (SEQ ID NO:1)). This peptide itself is immunologically inactive. The insertion of this peptide at the fusion point eliminates the neoantigenicity created by the joining of the two peptide moieties. The linker peptide also increases the flexibility of these moieties and allows retention of the biological activity. This relatively long linker peptide helps overcome the possible steric hindrance from the Fc portion of the hybrid, which could interfere with the activity of the hybrid.

The hybrid has a much longer half-life than the native IFN α . Due to the linker, it is also designed to reduce the possibility of generating a new immunogenic epitope (a neoantigen) at what would otherwise be the fusion point of the IFN α and the immunoglobulin Fc segment.

Cytokines are generally small proteins with relatively short half-lives which dissipate rapidly

among various tissues, including at undesired sites. It is believed that small quantities of some cytokines can cross the blood-brain barrier and enter the central nervous system, thereby causing severe neurological toxicity. The IFN α linked to Fc γ of the present invention would be especially suitable for treating hepatitis B or C, because these products will have a long retention time in the vasculature (upon intravenous administration) and will not penetrate undesired sites.

The specific hybrid described can also serve as a model for the design and construction of other cytokine-Fc hybrids. The same or a similar linker could be used in order to reduce the possibility of generating a new immunogenic epitope while allowing retention of the biological activity. Cytokine-Fc hybrids in which interleukin-2 is the cytokine, or hybrids including other cytokines, could be made using the same techniques.

Detailed Description of Making and Using the Invention

The hybrid molecule of the invention includes an interferon moiety linked through a unique linker to an immunoglobulin Fc moiety. Preferably, the C-terminal ends of two interferon moieties are separately attached to each of the two N-terminal ends of a heavy chain $\gamma 4$ Fc fragment, resulting in a homodimer structure. A unique linker peptide, Gly Gly Ser Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser (SEQ ID NO:1), was created to link the two moieties. The complete nucleotide sequence of the preferred $\gamma 4$ hybrid (including the linker and the Fc moiety) appears in SEQ ID NO: 7. The linker is located at amino acid residue numbers 189 to 204.

The advantage of the hybrid over the native cytokine is that the half-life *in vivo* is much longer. The hybrid including interferon and the $\gamma 4$ chain Fc homodimer is larger than the native interferon. Because the pores in the blood vessels of the liver are large, this larger molecule is more suitable for use in treating hepatitis, where the virus responsible primarily affects the liver.

The linker peptide is designed to increase the flexibility of the two moieties and thus maintain their biological activity. Although the interferon and the immunoglobulin are both of human origin, there is always a possibility of generating a new immunogenic epitope at the fusion point of the two molecules. Therefore, the other advantage of the linker of the invention, which consists mainly of a T cell inert sequence, is to reduce immunogenicity at the fusion point. Referring to SEQ ID NO:7, it can be seen that if the linker (residue numbers 189-204) was not present, a new sequence consisting of the residues immediately before number 189 and immediately after 204 would be created. This new sequence would be a neoantigen for the human body.

Human IFN α is derived from a family of several different genes. More than 24 species have been identified so far, from gene and protein sequence data. They differ from each other by anywhere from a few to a maximum of 35 amino acids. Most of the species have a signal peptide sequence of 23 amino acid residues and a mature amino acid sequence of 166 amino acid residues (Goeddel, D.V. et al., *Nature*, 290:20-26, 1981; Weissmann, C. and Weber, H., *Prog. Nuc. Acid Res. Mol. Biol.* 33:251-300, 1986; Zoon, K.C., *Interferon*, 9:1-12, 1987).

IFN α 2 (also called IFN α A) is one of the most intensively studied interferon species. The recombinant version of IFN α 2 has been used as a therapeutic for several years. Two IFN α 2 recombinant products, IFN α 2a and IFN α 2b, are now commercially available. They differ only in one amino acid at position 23, and there is no significant difference in biological activity between them (von Gabain, A., et al., *Eur. J. Biochem.* 190:257-61, 1990).

IFN α 2a was selected as the fusion partner for the interferon hybrid of the invention, although the IFN α 2b or any other interferon species (including IFN β) can be used as well. It is also possible to make similar constructs with other cytokines, such as interleukin-1 or interleukin-2. The same linker

into the culture supernatant where not many other proteins reside, making protein purification and process scale-up much easier. This system was tried first to express either the IFN α -Fc hybrid or the wild type IFN α 2a. Unfortunately the IFN α -Fc secreted was found to be partially degraded on SDS-PAGE, whereas the IFN α 2a alone was not. The degradation was believed to be caused by the protease activities present in the yeast expression system, as reported by Scorer, C.A. et al., *Gene*, 136:111-9, 1993. The relatively weak spot in the hinge region is the possible target for the proteases.

A mammalian cell expression system for the IFN α -Fc hybrid was also tried. The mammalian expression vector, pCDNA3 (Invitrogen, San Diego, CA) which contains a CMV promoter and a NEO resistance gene, was employed. The host cells, NSO cells, were transfected by the pCDNA3/IFN α -Fc expression vector using the electroporation method. The cells were selected by G418 at a concentration of 0.8 mg/ml. The IFN α -Fc expressing clones were identified by ELISA. The hybrid was successfully expressed in this system and there was no degradation.

There are several advantages to this mammalian expression system. First, the recombinant protein is secreted into the culture supernatant and there is no aggregation, thereby simplifying purification. One chromatography step using a protein A column yields a purified IFN α -Fc protein. Also, the protein produced in this system has a glycosylation pattern very similar to the natural molecules since it is expressed by mammalian cells. Further, a native IFN α 2a signal peptide sequence is included in the expression vector. Therefore the protein secreted from the cells has an authentic N-terminal, whereas in the *E. coli* or yeast expression systems there either is no signal peptide or a non-IFN α signal peptide is used. Either way, it will bring in additional artificial amino acid residue(s) at the N-terminal end of the recombinant IFN α -Fc.

As mentioned above, the purification of the IFN α -Fc recombinant protein from the culture

supernatant is relatively straightforward. The protein with a purity of more than 90%, as judged by SDS-PAGE, can be easily obtained by one step of affinity chromatography with a protein A column.

There are several assay methods available for the measuring of the IFN α bioactivity. Using an antiviral assay, it was demonstrated that the hybrid of SEQ ID NO:7 had a specific activity about 5 to 10 fold higher than a related IFN α -Fc hybrid, in which the linker molecule had the sequence Gly Gly Ser Gly Gly Ser (SEQ ID NO:2), and the Fc portion of the hybrid was derived from human IgG1 rather than IgG4. Nevertheless, although the biological activity of the hybrid shown in SEQ ID NO:7 was improved substantially over the similar hybrid, it was still lower than that of the native IFN α . However, it is expected that this hybrid will have a longer half-life *in vivo*, than the native IFN α . This expectation is based on results demonstrating that the related IFN α hybrid with the linker sequence shown in SEQ ID NO:2 and an IgG1 Fc portion showed a much longer half-life, in a pharmacokinetic study in a mouse model, than did the native IFN α .

Because the hybrid of SEQ ID NO:7 is expected to have a longer half-life *in vivo* than native IFN α , even though its specific activity is lower, this novel hybrid is expected to be preferred to the native IFN α for clinical use. This is because, as a result of the longer half-life, the Cxt (the area under the concentration vs. time curve) would be up to several hundred times greater than for the native IFN α . This means that at the equivalent molar dosage of the native IFN α and the hybrid, the latter would provide a several hundred fold increased exposure to IFN α , resulting in vastly increased efficacy at the same dosage, and less frequent administration.

20 In measuring specific activity, molar dosage is preferred instead of expressing activity as units per mass of protein. This is because interferons function through the binding to their specific receptors, which is directly related to the number of molecules present. Also, the molecular weight

of the IFN α -Fc γ 4, 110 Kd, is more than five-fold larger than that of the wild type IFN α 2a, which is 20kd. Taking this into consideration, measuring activity in units/ μ mol instead of the units/mg provides a better comparison of activity specificity.

Example I: Cloning human IFN α cDNA and constructing the IFN α -Fc expression vector

5 6×10^6 KG-1 cells (ATCC 246) were incubated with 200 units of Sendai virus at 37°C overnight. The cells were harvested and washed with PBS thoroughly. The total RNA was extracted by using the RNA-ZOL RNA isolation kit (BIOTEX, Houston, TX) following the procedure provided by the manufacturer. The first-strand cDNA was synthesized by reverse transcription using AMV reverse transcriptase with oligo(dT) as 3' primer in 50mM Tris-HCl (pH 8.3), 60mMKCl, and 6mM MgCl₂, incubated at 42°C for 1 hour. The reaction mixture was used directly as the template for PCR to amplify IFN α cDNA. The 5' primer for PCR contained a Hind III site and the coding sequence for the first 21 amino acids from the IFN α 2a leader peptide (SEQ ID NO:3). The 3' primer contained the sequence coding for part of the linker (SEQ ID NO:1) and the last five amino acids of the IFN α 2a, and a BamH I site integrated in the linker sequence (SEQ ID NO:4). The PCR buffer contained 50mM KCl, 10mMTris-Hcl (pH8.3), 1.5mM MgCl₂, 0.01% gelatin, 0.1 mmol each of dNTP, 0.5 μ mol of each primers, 5 μ l RT reaction mixture, and 1 unit of Taq DNA polymerase in a total of 50 μ l volume. The PCR condition was 94°C (1 min), 55°C (2 min), and 72°C (2 min) for 40 cycles on a GeneAmp PCR System 9600 (Perkin Elmer, Norwalk, CT).

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The cDNA of the human immunoglobulin γ 4 Fc was obtained by reverse transcription and PCR performed the same way as described above. The RNA was extracted from the human tonsil B cells. The 5' primer had the sequence shown in SEQ ID NO:5. The 3' primer had the sequence shown in SEQ ID NO:6.

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The two PCR amplified DNA segments were cloned into pUC18 vectors at sites Hind III/BamH I or sites BamH II/EcoR I respectively. After their DNA sequences were confirmed by DNA sequencing using the kit from USB (Cleveland, Ohio), the two segments were ligated together through the BamH I site by a second round cloning. The full length IFN α -Fc cDNA was then inserted into a mammalian expression vector pCDNA3 (Invitrogen, San Diego, CA) through the Hind III and EcoR I sites.

Example 2: Expressing IFN α -Fc in mammalian cells

10⁷ NSO cells were mixed with 10 μ g linearized pCDNA3/IFN α -Fc plasmid in 0.8 ml PBS and kept on ice for 5 min. Electroporation was performed at 200v, 960 μ F using Gene Pulser (BioRad, Hercules, CA). The cells were then put back on ice for 20 minutes and transferred to a 100mm tissue culture plate in 10ml DMEM supplied with 2% FCS. After incubation at 37°C for two days, the cells were washed and resuspended in the same medium. 0.6 mg/ml G418 was added to start the selection. The cells were plated out in eight 96-well micro plates and incubated at 37°C. Colonies appeared in one week and they were ready for screening in two weeks. The supernatants from each well with a single colony growing were collected. The IFN α -Fc in the supernatant was quantitatively determined by an ELISA assay employing goat anti-human IgG and anti-human Fc conjugated with horseradish peroxidase. The clones with higher ELISA readings and smaller colony size were selected for subcloning. These colonies were transferred to a 24-well plate and supplied with a medium containing G418. The clone with the highest secretion level was expanded and adapted to grow in a spinner. For large scale preparation, the culture supernatant was collected and passed through a protein A agarose column equilibrated by PBS. The protein bound to the protein A was eluted by 50 mM citric acid (pH 3.0) and concentrated by lyophilization.

Example 3: Characterization of the IFN α -Fc hybrid.

The purity of the recombinant protein isolated from NSO culture medium was examined by SDS-PAGE and Western blot. Only one protein band was visible on the blotted membrane stained by ponceau s for total proteins, showing a homogeneity of the protein preparation. The apparent molecular weight of this protein is about 55kd under reducing conditions and 110kd under non-reducing conditions, which is exactly the predicted size for the IFN α -Fc hybrid. The doubling of its apparent molecular weight under non-reducing conditions suggests that the hybrid is in a dimeric form. The recombinant protein can be recognized by both anti-Fc and anti-IFN α antibodies, confirming that it consists of two moieties, the IFN α and the Fc fragment.

The bioactivity assay for the IFN α -Fc was an antiviral assay. Specifically, the assay method used was a modification of the protocol described by Robert M. Friedman et al (Measurement of antiviral activity induced by interferons α , β , and γ , *Current Protocols in Immunology*, 1994, pp. 6.9.1-6.9.8). Briefly, human lung carcinoma cells (A549, ATCC#CCL 185) were seeded in 96-well plates at a density of 40,000 cells/well and incubated at 37°C for 24 hours. 1:2 serially diluted IFN α -Fc hybrid or native IFN α (NIH# Gxa01-901-535) were added and incubated at 37°C for 24 hours. Every sample was done in triplicate. The culture medium was replaced with a fresh one containing encephalomyocarditis virus (ATCC #VR 1298) at a concentration of about 0.1 MOI/cell and incubated at 37°C for a further 48 hours. The dead cells were washed away by pipetting up and down vigorously with PBS. The attached cells were fixed by 2% formaldehyde and stained by giemsa stain. The plates were rinsed with tap water and allowed to dry. The stained cells were dissolved by methanol and the samples were read spectrophotometrically at 595nm. The antiviral activity of IFN α -Fc hybrid was calculated by comparing it with the IFN α standard, and was found to be about 30 to 60% of the activity of the IFN α standard.

It should be understood that the terms and expressions used herein are exemplary only and not limiting, and that the scope of the invention is defined only in the claims which follow, and includes all equivalents of the subject matter of those claims.

SEQUENCE LISTING

(1) General Information:
(i) Applicant: Yu, Liming; Chang, Tse Wen
(ii) Title of Invention: Hybrid with Interferon- α and an Immunoglobulin Fc Linked through a Non-Immunogenic Peptide
5 (iii) Number of Sequences: 7
(iv) Correspondence Address:
(A) Addressee: Tanox Biosystems, Inc.
(B) Street: 10301 Stella Link Rd.
10 (C) City: Houston
(D) State: Texas
(E) Country: USA
(F) Zip: 77025
(v) Computer Readable Form:
(A) Medium Type: Diskette, 3.5 inch
(B) Computer: Addonics C142 SVGA
(C) Operating System: DOS 3.30
(D) Software: Wordperfect 5.1
15 (vi) Current application data:
(A) Application Number:
(B) Filing Date:
(C) Classification:
(vii) Prior Application Data:
(A) Application Number: 08/579,211
20 (B) Filing Date: 12/28/95
(viii) Attorney/Agent Information:
(A) Name: Mirabel, Eric P.
(B) Registration Number: 31,211
25 (C) Reference/Docket Number: 95-2-PCT
(ix) Telecommunication Information:
(A) Telephone: (713) 664-2288
(B) Telefax: (713) 664-8914
(2) Information for SEQ ID NO:1:
30 (i) Sequence Characteristics:
(A) Length: 48 nucleic acids
(B) Type: nucleotide
(C) Strandedness: double stranded
(D) Topology: linear
35 (xi) Sequence Description: SEQ ID NO:1:
40

GGT GGC TCA GGT GGA TCC GGT GGA GGC GGA AGC GGC 36
Gly Gly Ser Gly Gly Ser Gly Gly Gly Ser Gly
1 5 10

45 GGT GGA GGA TCA 48
Gly Gly Gly Ser
15

50 (2) Information for SEQ ID NO:2:
(i) Sequence Characteristics:
(A) Length: 6 amino acids
(B) Type: amino acid
(D) Topology: unknown
55 (xi) Sequence Description: SEQ ID NO:2:

Gly Gly Ser Gly Gly Ser
1 5

60 (2) Information for SEQ ID NO:3:
(i) Sequence Characteristics:

GAC ACT CTC ATG ATC TCC CCG ACC CCT GAG GTC ACG TGC 741
 Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
 235 240 245
 5 GTG GTG GTG GAC GTG AGC CAG GAA GAC CCC GAG GTC CAG 780
 Val Val Val Asp Val Ser Gln Glu Asp Pro Glu Val Gln
 250 255 260
 10 TTC AAC TGG TAC GTG GAT GGC GTG GAG GTG CAT AAT GCC 819
 Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala
 265 270
 15 AAG ACA AAG CCG CGG GAG GAG CAG TTC AAC AGC ACG TAC 858
 Lys Thr Lys Pro Arg Glu Glu Gln Phe Asn Ser Thr Tyr
 275 280 285
 20 CGT GTG GTC AGC GTC CTC ACC GTC CTG CAC CAG GAC TGG 897
 Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp
 290 295
 25 CTG AAC GGC AAG GAG TAC AAG TGC AAG GTC TCC AAC AAA 936
 Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
 300 305 310
 30 GGC CTC CCG TCC TCC ATC GAG AAA ACC ATC TCC AAA GCC 975
 Gly Leu Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys Ala
 315 320 325
 35 AAA GGG CAG CCC CGA GAG CCA CAG GTG TAC ACC CTG CCC 1014
 Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro
 330 335
 40 CCA TCC CAG GAG GAG ATG ACC AAG AAC CAG GTC AGC CTG 1053
 Pro Ser Gln Glu Glu Met Thr Lys Asn Gln Val Ser Leu
 340 345 350
 ACC TGC CTG GTC AAA GGC TTC TAC CCC AGC GAC ATC GCC 1092
 Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala
 355 360
 45 GTG GAG TGG GAG AGC AAT GGG CAG CCG GAG AAC AAC TAC 1131
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 365 370 375
 50 AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GGC TCC TTC 1170
 Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe
 380 385 390
 55 TTC CTC TAC AGC AGG CTA ACC GTG GAC AAG AGC AGG TGG 1209
 Phe Lys Tyr Ser Arg Leu Thr Val Asp Lys Ser Arg Trp
 395 400
 60 CAG GAG GGG AAT GTC TTC TCA TGC TCC GTG ATG CAT GAG 1248
 Gln Glu Gly Asn Val Phe Ser Cys Ser Val Met His Glu
 405 410 415
 GCT CTG CAC AAC CAC TAC ACA CAG AAG AGC CTC TCC CTG 1287
 Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu
 420 425

TCT CTG GGT AAA TAG 1302
Ser Leu Gly Lys
430

What Is Claimed Is:

1. A hybrid molecule comprising an interferon molecule joined at its C-terminal end through a peptide linker to the N-terminal end of the immunoglobulin Fc fragment, the peptide linker comprising the sequence Gly Gly Ser Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser (SEQ ID NO:1).
5
2. The hybrid molecule of claim 1 in which another interferon molecule is joined at its C-terminal end through the peptide linker to the N-terminal end of a chain of the immunoglobulin Fc fragment, thereby forming a homodimer.
3. The hybrid molecule of claim 2 in which the interferon molecule is IFN α 2a
10 or IFN α 2b.
4. The hybrid molecule of claim 2 in which the Fc fragment is a γ 4 chain Fc fragment.
5. A method of treating hepatitis, hairy cell leukemia, multiple myeloma, or other cancers or viral diseases, comprising administering the hybrid molecule of any of
15 claims 1 to 4.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US96/20861

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : A61K 38/21, 39/395, 39/44; C07K 14/56
 US CL : 424/134.1; 435/69.7; 514/2, 12, 934; 530/351, 387.3; 536/23.52
 According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/134.1; 435/69.7; 514/2, 12, 934; 530/351, 387.3; 536/23.52

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
 NONE

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
 APS, DIALOG, MEDLINE, EMBASE, BIOSIS, WORLD PATENT INDEX

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,349,053 A (LANDOLFI) 20 September 1994, columns 4 and 13.	1-5
Y	WO 91/16353 A1 (CORVAS INTERNATIONAL N.V.) 31 October 1991, figure 6, sequence 9, pages 16-20.	1-5
Y	BARON, E. et al. From Cloning to a Commercial Realization: Human Alpha Interferon. Biotechnology. 1990. Vol. 10. No. 3. pages 197-190, see especially page 185.	1-5

Further documents are listed in the continuation of Box C. See patent family annex.

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Date of the actual completion of the international search 14 FEBRUARY 1997	Date of mailing of the international search report 04 APR 1997
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